EtOAc-Et₂O, yield 4.30 g, and combined with 5.40 g from preparation A and recrystallized from EtOAc: yield 7.9 g; mp 188-190 °C; [α]²⁵D -34.8° (1% in 95% EtOH); M⁺ 358; UV λ_{max} 238 nm (ϵ 12800); NMR δ 0.88 (3, C-18 CH₃), 1.33 (3 C-19 CH₃), 3.29 (1, ==CH), 3.60 (2, OCH₂), 4.40 (1, HOCH₂), 5.07 (1, 6β-OH), 5.27 (1, 17β-OH), 5.67 (1, C₄==CH). Anal. (C₂₂H₃₀O₄) C, H. Treatment of 8 with Ac₂O-Py afforded the diacetate 9 which

was used without purification for NMR (see discourse). 6β , 17-Dihydroxy-2-(hydroxymethyl)-17 α -pregna-1,4-

dien-20-yn-3-one (10). Organism Melanospora parasitica (ATCC 11 103) (Me₁): five runs of 2.0 g of 7 for 48 h; crude product from EtOAc; yield 4.5 g; mp 226-227 °C; $[\alpha]^{25}D$ -81° (1% in 95% EtOH); M⁺ 356; UV λ_{max} 250 nm (ϵ 15800); NMR δ 0.87 (3, C-18 CH₃), 1.41 (3, C-19 CH₃), 3.23 (1, C=C-H), 4.20 (2, OCH₂), 4.40 (1, CHO), 4.85 (1, CH₂OH), 5.21 (1, CHOH), 5.30 (1, C-17 OH), 6.03 (1, ==CH), 7.07 (1, ==CH). Anal. (C₂₂H₂₈O₄) C, H.

Metabolite Isolation. The following TLC systems were utilized in metabolite studies using silica gel PF_{254} plates: (1) EtOAc; (2) EtOAc-cyclohexane (1:1); (3) $CH_2Cl_2-CH_3OH$ (9:1); (4) EtOAc-cyclohexane-isopropylamine (25:24:1); (5) EtOAc-MeOH (9:1); (6) Et_2O ; (7) EtOAc-MeOH (7:3); (8) EtOAc-*n*-hexane (1:1).

Aliquots of crude extracts of urine were applied in narrow bands on TLC plates and developed in one of the above solvent systems. Components were visualized under 253-nm UV light or by spraying sections of the plate with 50% EtOH- H_2SO_4 and heating for color development. Bands were removed and the silica gel was placed in a glass micro column and eluted with MeOH. After evaporation the material was put on another plate which was developed in a second solvent system. Isolated metabolites were compared with reference compounds in several solvent systems.

Monkey Study. A 24-h urine sample was collected from three female Rhesus monkeys medicated orally with danazol in Tween 80 at 50 mg/kg b.i.d. for a total of 664 mg. A 100-ml sample was extracted with *n*-hexane (2×200 ml) which was discarded. The sample was then extracted with CH₂Cl₂ (2×200 ml) which was dried (Na₂SO₄) and evaporated and the residue purified by TLC with system 1 and then system 8. Several metabolites were observed but only one, ethisterone (11), was obtained in sufficient purity to allow identification (Table I).

Human Study. Urine from a female subject administered 800 mg of danazol orally was collected for 24 h. A 1.8-l. aliquot was extracted with CH_2Cl_2 (2 × 4 l.). After being washed with 1 l. of 0.1 N NaOH, 1 l. of 0.1 N HCl, and 1 l. of water, the CH_2Cl_2 solution was dried (Na₂SO₄) and the solvent removed by distillation. The residue was chromatographed using system 3 to give five components which were rechromatographed and the products isolated and identified: metabolite 1, purified in system 2, was identical with ethisterone (11); metabolite 2, purified in system 6, was identical with 7; metabolite 4, purified in system 5, was

identical with 8; metabolite 5, purified in system 7, was identical with 10. See Table II.

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References and Notes

- A. J. Manson, F. W. Stonner, H. C. Neumann, R. G. Christiansen, R. L. Clarke, J. H. Ackerman, D. F. Page, J. W. Dean, D. K. Phillips, G. O. Potts, A. Arnold, A. L. Beyler, and R. O. Clinton, J. Med. Chem., 6, 1 (1963).
- (2) G. O. Potts, A. Beyler, and H. P. Schane, Fertil. Steril., 25, 367 (1974).
- (3) (a) R. B. Greenblatt, W. P. Dmowski, V. B. Mahesh, and H. F. L. Scholer, *Fertil. Steril.*, 22, 102 (1971); (b) R. L. Friedlander, J. Reprod. Med., 10, 197 (1973); (c) R. D. Skoglund and C. A. Paulsen, Contraception, 7, 357 (1973); (d) R. J. Sherins, H. M. Gandy, T. W. Thorslund, and C. A. Paulsen, J. Clin. Endocrinol. Metab., 32, 522 (1971); (e) R. B. Greenblatt, M. I. Oettinger, R. Borenstein, and C. S. S. Bohler, J. Reprod. Med., B 201 (1974); (f) M. C. Andrews and A. C. Wentz, Am. J. Obstet. Gynecol., 121, 817 (1975); (g) W. P. Dmowski and M. R. Cohen, Obstet. Gynecol., 46, 147 (1975); (h) R. B. Greenblatt, R. Borenstein, and S. Hernandez-Ayup, Am. J. Obstet. Gynecol., 118, 783 (1974).
- (4) The hormones, their current status, Sixth Drug Trade News Survey reprinted from Sept 3 and Sept 17, 1962 issues of Drug Trade News, p XII.
- (5) T. Takegoshi, H. Tachizawa, and G. Ohta, Chem. Pharm. Bull., 20, 1243 (1972).
- (6) C. Davison, W. Banks, and A. Fritz, Arch. Int. Pharmacodyn. Ther., 222, 294 (1976).
- (7) A. J. Manson, R. E. Sjogren, and M. Riano, J. Org. Chem., 30, 307 (1965).
- (8) E. J. Corey and D. E. Cane, J. Org. Chem., 36, 3070 (1971).
- (9) J. A. Edwards, M. C. Calzada, L. C. Ibánez, M. F. Cabezas Rivera, R. Urquiza, L. Cardona, J. C. Orr, and A. Bowers, J. Org. Chem., 29, 3481 (1964).
- (10) L. Smith, Steroids, 1, 570 (1963).
- (11) A. Cohen and S. Rock, Jr., Steroids, 3, 243 (1964).
- (12) C. Gerali, G. C. Sportoletti, C. Parini, A. Ius, and L. Cecchi, *Farmaco, Ed. Sci.*, 24, 306 (1969).
- (13) N. M. Yoon and H. C. Brown, J. Am. Chem. Soc., 90, 2927 (1968).

Ring D Oxygenated Spirolactones. Characterization of a Human Metabolic Product of Spironolactone

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 15α -Hydroxycanrenone (1b) was prepared from canrenone (1a) by microbiological oxidation with a *penicillium* species. The product was identical with one obtained from the metabolism of spironolactone (3) in human. Oxidation of 1b with Jones reagent furnished the corresponding 15-oxocanrenone (1d) which underwent base-catalyzed β elimination to generate an α,β -unsaturated cyclopentenone system. 15α -Hydroxycanrenone (1b) failed to show antimineralocorticoid activity at the screening dose of 2.4 mg while the oxo derivative 1d exhibited approximately 15% the activity of 3. Since the activity of canrenone is 38% that of spironolactone, introduction of the carbonyl group at the 15 position of canrenone resulted in a reduction in activity. This effect is opposite to that observed with 6-dehydroprogesterone.

In earlier studies on the antimineralocorticoid activity of various progesterone derivatives, the effect which an oxygen function at the 15 position had on this activity was examined.¹ In extending these studies, we sought to introduce an oxygen function into C-15 of $3-(17\beta-hy-droxy-3-oxoandrosta-4,6-dien-17\alpha-yl)$ propionic acid γ -lactone (canrenone, 1a) in order to determine whether it would alter the mineralocorticoid-blocking activity of this

substance.

Using a *penicillium* species (our code number M31-417) which had previously been found to hydroxylate androst-4-ene-3,17-dione at the 15α position in our laboratory (unpublished results), we observed that 1a was converted into a hydroxylated derivative, presumably 15α -hydroxvcanrenone (1b). A structure determination of this substance was undertaken because 2a, the 6,7-dihydro derivative of canrenone, had previously been found to be hydroxylated by a penicillium species (M31-399) to afford the 15 β -hydroxylated derivative **2b** rather than the epimeric compound $2c^2$. The latter product was, however, obtained by hydroxylation with a cephalosporium species (M60-20)² Subsequently 1b was found to be identical with a substance obtained from the metabolic study of spironolactone 3 in human,³ and this made an unambiguous assignment of its structure even more crucial.

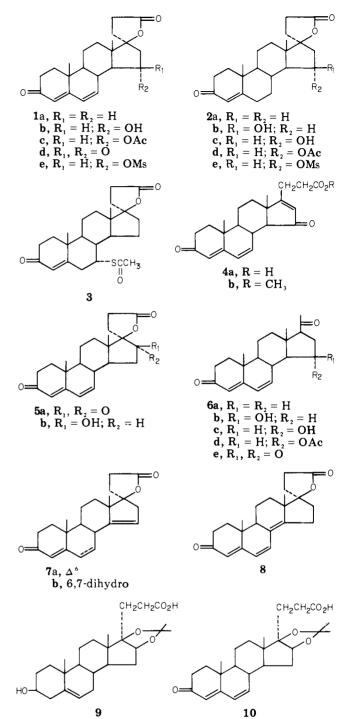
Although 1b was obtained from the fermentation mixture as an amorphous substance, even after chromatography, it formed a crystalline pyridine complex. Conversion to the acetate 1c and oxidation to give the ketone 1d established that the newly introduced hydroxyl group was secondary. The carbonyl stretching frequency of 1745 cm⁻¹ revealed that the saturated keto group of 1d was located in a five-membered ring.⁴ Thus, hydroxylation occurred at either C-15 or C-16. Treatment of the ketone 1d with methanolic sodium methoxide resulted in β elimination to afford the unsaturated keto acid 4a. The presence of the cyclopentenone chromophore and the 3-keto- $\Delta^{4,6}$ system was demonstrated by ultraviolet maxima at 230-232 and 286-289 nm, respectively. Comparable values were obtained for the corresponding methyl ester 4b. The results indicated that hydroxylation had occurred at C-15.

Further evidence that C-16 was not the point of attack in the hydroxylation process came from a comparison of the mass spectrum of the ketone 1d with that of 16oxocanrenone 5a. The latter substance was prepared from $3 \cdot (3\beta, 16\beta, 17\beta$ -trihydroxyandrost-5-en- 17α -yl)propionic acid 16,17-acetonide (9).⁵ Conversion of the 3β -hydroxy- Δ^5 system of 9 into the 3-keto- $\Delta^{4,6}$ -diene system of 10 was accomplished by means of a mixture of bromine, lithium bromide, and lithium carbonate in dimethylformamide.⁶ Removal of the acetonide-protecting group followed by lactonization afforded 5b. Oxidation of the 16β -hydroxyl group gave the ketone 5a. The mass spectra of 5a and 1d were found to be dissimilar.

The transformation of canrenone 1a into the 15-hydroxy derivative 1b resulted in a dextrorotatory shift of the molecular rotation ($\Delta M_{\rm D} = 132$). Acetylation or oxidation of the 15-hydroxyl group produced a levorotatory shift ($\Delta M_{\rm D} = -20$ and -65, respectively). This pattern was observed also in the 6-dehydroprogesterone series in which the hydroxyl group at C-15 is known to be α oriented.^{1a} Where the hydroxyl group is 15 β , the molecular rotational difference between 6-dehydroprogesterone (6a) and the 15-hydroxy derivative 6b was found to be rather slight ($\Delta M_{\rm D} = 6$).

The chemical shifts of the 6 and 7 protons are coincident in both canrenone (1a) and 6-dehydroprogesterone (6a).^{1a} This is true also for the corresponding 15α -acetoxy derivatives 1c and 6d. However, in the 15α -hydroxy (1b and 6c) and 15-oxo (1d and 6e) derivatives, the chemical shifts of the 6 and 7 protons are different, as they are also in 15β -hydroxy-6-dehydroprogesterone (6b).

Dehydrogenation of 2d with dichlorodicyanobenzoquinone in the presence of hydrogen chloride furnished a product which was identical with the $\Delta^{4,6}$ compound 1c.



This provided further evidence that the product obtained from hydroxylation of canrenone (1a) with *penicillium* sp. M31-417 was, indeed, 15α -hydroxycanrenone (1b).

In both the canrenone and the dihydrocanrenone series, elimination of the 15α -hydroxy group to give the corresponding Δ^{14} derivatives, **7a** and **7b**, was achieved by conversion first to the mesylates 1e and 2e, followed by treatment with either potassium or sodium acetate. Dehydrogenation with chloranil converted **7b** into **7a**. Isomerization of the latter substance in an alkaline medium furnished the conjugated trienone 8.

The antimineralocorticoid effects of the various derivatives of canrenone, in which ring D has been modified, were determined in a standard test employing deoxycorticosterone acetate as the mineralocorticoid. The compounds were administered subcutaneously at a dose of 2.4 mg per rat, each rat weighing between 175 and 200 g. A compound was considered active if the increase in

Table I.Antimineralocorticoid Effects of Ring DModified Spirolactones

Compd	Urinary log (Na \times 10)/K		
	$\frac{\text{Treated}}{(T)^{a}}$	Control (C) ^b	T/C^c
1b	0.57	0.91	0.62
1 c	0.77	0.99	0.77
1 d	1.04	1.03	1.01
5b	0.97	1,16	0.84
7 a	0.55	0.82	0.67
8	0.73	0.89	0.82

^a Each of the compounds was tested in the adrenalectomized rat at 2.4 mg subcutaneously following the subcutaneous administration of 12 μ g of deoxycorticosterone acetate. Eight animals were employed in each group. ^b Spironolactone was given to the adrenalectomized rat at 0.33 mg subcutaneously following the subcutaneous administration of 12 μ g of deoxycorticosterone acetate. Groups of eight animals were employed. ^c A compound is considered active when T/C \geq 1.

the urinary log (Na \times 10)/K ratio was equivalent to or greater than that produced by 0.33 mg/rat of spironolactone [3-(3-oxo-7 α -acetylthio-17 β -hydroxyandrost-4en-17 α -yl)propionic acid lactone (3)].^{5,7}

With the exception of 15-oxocanrenone (1d), all of the derivatives examined were found inactive in this test at the screening dose (Table I). The activity of 15-oxocanrenone (1d) was approximately 15% that of spironolactone (3). The activity of canrenone (1a), on the other hand, has been reported to be ca. 35% that of spironolactone when administered orally.⁸ Given subcutaneously, the activity of 1a is 38% that of $3.^9$

Significantly, introduction of a carbonyl group into the 15 position of canrenone (1a) did not lead to enhancement of antimineralocorticoid activity. This is in contrast to the results obtained with 6-dehydroprogesterone (6a) wherein insertion of the 15-carbonyl group led to increased activity.^{1a}

Earlier we had observed that a 6β , 7β -methylene group enhanced the antimineralocorticoid effect of a spirolactone derivative while the same group had the opposite effect in the progesterone series.^{1b,10} These results indicate that the structural features required for antimineralocorticoid activity differ in the two series of compounds which are known to block the salt-retaining effects of aldosterone.

Experimental Section

Melting points were determined on a Fisher-Johns melting block and are uncorrected. NMR spectra were taken on a Varian A-60 instrument in deuteriochloroform with tetramethylsilane as an internal standard. Optical rotations were determined in CHCl₃.

3-(15 α ,17 β -Dihydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic Acid γ -Lactone (1b). A 12.0-g sample of 3-(17 β hydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic acid γ -lactone (1a) in 25 l. of cotton seed meal broth was fermented in a 40-l. fermentor for a period of 48 h with *penicillium* sp. M31-417 and worked up in the usual manner.¹¹ The crude reaction product was chromatographed on 1.2 kg of silica gel. The column was eluted with varying proportions of C₆H₆ and EtOAc. Elution with 50% EtOAc in C₆H₆ gave 2.8 g of 1b as an amorphous substance. A 698-mg sample of 1b was crystallized from pyridine to afford 407 mg of 1b as a pyridine complex: mp 112–117 °C; NMR (Hz) 431–472 (multiplet, C₅H₅N protons), 400 (dd, J = 10, 1.5 Hz, 1 H, C₇-H), 366 (dd, J = 10, 2.5 Hz, 1 H, C₆-H). Anal. (C₂₂H₂₈-O₄·C₅H₅N) C, H, N.

A sample of the pyridine complex of **1b** was dissolved in EtOAc. The EtOAc solution was evaporated to dryness to afford a viscous oil. The oil was triturated with ether to afford **1b**, as an amorphous solid: mp 120–125 °C; $[\alpha]^{25}$ D 60.3° (c 1); λ_{max}^{MeOH} 281–283.5 nm (ϵ 24 300); c (KBr) 3695, 3620, 3470, 1770, 1660, 1654, 1621 cm⁻¹;

NMR (Hz) 398 (q, J = 10, 2 Hz, 1 H, C₇-H), 367 (q, J = 10, 2 Hz, 1 H, C₆-H). The NMR spectrum showed the absence of pyridine.

3-(15 α -Acetoxy-17 β -hydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic Acid γ -Lactone (1c). A mixture of 326 mg of 1b, 1 ml of C₅H₅N, and 1 ml of Ac₂O was heated on the steam bath for 1 h. The cooled reaction mixture was poured into ice water and stirred. The resultant solid was collected, washed with water, and dried. The solid was extracted with boiling ether. The ether extract was concentrated to afford a crystalline product which was recrystallized from ether to furnish 1c: mp 208–212 °C; [α]²⁵D 49° (c 0.1); ν (KBr) 1778, 1747, 1674, 1631, 1595 cm⁻¹; NMR (Hz) 368 (s, 2 H, C₆,C₇-H's), 343 (s, 1 H, C₄-H), 124.5 [s, 3 H, CH₃C(==O)O-], 69 (s, C₁₀-CH₃), 65.5 (s, C₁₃-CH₃). Anal. (C₂₄-H₃₀O₅) C, H.

3-(17β-Hydroxy-3,15-dioxoandrosta-4,6-dien-17α-yl)propionic Acid γ-Lactone (1d). A stirred solution of 350 mg of 1b in 7 ml of acetone was treated portionwise with a solution of 0.35 ml of Jones reagent (8 N CrO₃) in 2 ml of acetone. The reaction mixture was stirred at room temperature for 20 min after which it was diluted with H₂O. The resultant precipitate was collected and dried. Crystallization from acetone afforded 202 mg of 1d: mp 218-219 °C; ν (KBr) 1795, 1745, 1675, 1628, 1593 cm⁻¹; λ_{max} ^{MeOH} 280-282 nm (ϵ 26000); NMR (Hz) 412 (dd, J =10, 2 Hz, 1 H, C₇-H), 370 (dd, J = 10, 2.5 Hz, 1 H, C₆-H), 342.5 (1 H, C₄-H), 69.5 (s, 3 H, C₁₀-CH₃), 68.5 (s, 3 H, C₁₃-CH₃). Anal. (C₂₂H₂₆O₄) C, H.

3-(15 α ,17 β -Dihydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic Acid γ -Lactone 15-Methanesulfonate (1e). To a mixture of 100 mg of the pyridine complex of 1b and 0.5 ml of pyridine was added 0.1 ml of methanesulfonyl chloride. The reaction mixture was allowed to stand at room temperature for 1 h with occasional swirling. Then it was allowed to stand at 5 °C for 15 h. The reaction mixture was diluted with H₂O and stirred. The precipitate which resulted was collected, washed with H₂O, and dried. Crystallization from EtOAc-ether afforded 1e, mp 135-140 °C. The presence of a sharp signal at 297.5 Hz (3-H) in the NMR spectrum confirmed the presence of the methanesulfonyl group in 1e.

3-(3,15-Dioxoandrosta-4,6,16-trien-17-yl)propionic Acid (4a). A mixture of 103 mg of the 15-oxospirolactone (1d), 10 ml of MeOH, and 60 mg of NaOMe was stirred at room temperature in an atmosphere of N₂ for 10 min. The reaction mixture was diluted with H₂O and acidified with glacial HOAc. The resultant mixture was concentrated under reduced pressure until a heavy oil appeared. The oil was extracted with EtOAc. The EtOAc extract was washed with H₂O, dried (Na₂SO₄), and evaporated to dryness to afford 118 mg of a yellow foam: λ_{max}^{MeOH} 230–232 nm (ϵ 12900), 286–289 (23100); ν (KBr) 1730, 1708, 1666, 1640, 1618 cm⁴. Crystallization of the foam from CH₂Cl₂-C₆H₆ afforded 4a: mp 198–200 °C; NMR (Hz, after D₂O exchange) 389 (dd, J = 10, 1.5 Hz, 1 H, C₆ or C₇-H), 370 (dd, J = 10, 2.5 Hz, 1 H, C₇ or C₆-H), 351 (s, 1 H, C₁₆-H), 342 (s, 1 H, C₄-H).

Methyl 3-(3,15-Dioxoandrosta-4,6,16-trien-17-yl)propionate (4b). A mixture of 75 mg of 4a and 15 ml of an ethereal solution of CH₂N₂ prepared from 700 mg of N-nitrosomethylurea was allowed to stand at 5 °C for 66 h. The reaction mixture was evaporated to dryness. The residual oil was chromatographed on 3.0 g of silica gel. The column was eluted with varying proportions of EtOAc in C₆H₆. Elution with 15% EtOAc in C₆H₆ afforded 4b as a yellow viscous oil: λ_{max}^{MeOH} 227.5-231 nm (ϵ 13 200), 287 (ϵ 24 800). The appearance of a signal at 222 Hz in the NMR spectrum confirmed the presence of the carbomethoxy group.

3- $(15\alpha$ -Acetoxy- 17β -hydroxy-3-oxoandrost-4-en- 17α -yl)propionic Acid γ -Lactone (2d). A mixture of 2.0 g of 3- $(15\alpha, 17\beta$ -dihydroxy-3-oxoandrost-4-en- 17α -yl)propionic acid γ -lactone (2c),² 10 ml of C₅H₅N, and 6.4 ml of Ac₂O was maintained at room temperature for 20 h after which it was poured into ice H₂O. The resultant precipitate was collected, washed with H₂O, and dried: yield 2.05 g. Crystallization from EtOAc afforded 2d: mp 186–192 °C; NMR (Hz) 345 (s, 1 H, C₄-H), 73 (s, 3 H, C₁₀-CH₃), 63 (s, 3 H, C₁₃-CH₃). Anal. (C₂₄H₃₂O₅) C, H.

Dehydrogenation of $3 \cdot (15\alpha - \text{Acetoxy} \cdot 17\beta - \text{hydroxy} \cdot 3 \cdot \text{oxo-androst} \cdot 4 \cdot \text{en} \cdot 17\alpha \cdot \text{yl})$ propionic Acid γ -Lactone (2d). A mixture of 100 mg of 2d, 60 mg of dichlorodicyanobenzoquinone, and 7 ml of 5.9 N HCl in dioxane was stirred and heated under reflux

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for 45 min.¹² Then it was allowed to stand at room temperature for 15 h. The reaction mixture was diluted with ether and 5% NaOH. The organic phase was separated, washed successively with 5% NaOH and H₂O, dried (Na₂SO₄), and evaporated to dryness to afford a yellow viscous oil. Crystallization from ether afforded 65 mg of 1c: mp 208–212 °C; λ_{max}^{MeOH} 279–281 nm (ϵ 26 200). IR and NMR spectroscopy, as well as GLC, confirmed that the product was identical with that obtained from the acetylation of 1b.

3- $(17\beta$ -Hydroxy-3-oxoandrosta-4,6,14-trien- 17α -yl)propionic Acid γ -Lactone (7a). (a) A mixture of 100 mg of methanesulfonate le, 100 mg of fused KOAc, and 8 ml of glacial HOAc was heated under reflux for 1.5 h. Then it was distilled nearly to dryness under reduced pressure. The residue was triturated with H₂O. The resultant solid was collected, washed with H₂O, and dried. Crystallization from EtOAc-ether-hexane afforded 39 mg of 7a, mp 215-220 °C.

(b) A solution of 5.0 g of $3-(15\alpha,17\beta$ -dihydroxy-3-oxoandrost-4-en-17 α -yl)propionic acid γ -lactone (2c) in 10 ml of C_5H_5N was cooled in an ice bath and stirred while 3.0 ml of methanesulfonyl chloride was added dropwise over a period of 12 min. Stirring was continued while the reaction mixture was allowed to warm to room temperature over a period of 75 min. The reaction mixture was poured into ice H_2O . The precipitate was collected, washed with H_2O , and dried. The solid was digested with 12 ml of hot EtOH. On cooling, the EtOH solution afforded the mesylate 2e as a semisolid.

A 3.9-g sample of **2e**, 1.95 g of freshly fused NaOAc, and 12 ml of glacial HOAc was heated under reflux in an atmosphere of N₂ for 25 min. The cooled reaction mixture was diluted with H₂O. The precipitate was collected, washed with H₂O, and dried. Crystallization from MeOH afforded 1.0 g of 3-(17 β -hydroxy-3-oxoandrosta-4,14-dien-17 α -yl)propionic acid γ -lactone (**7b**): mp 188–191 °C; NMR (Hz) 346 (s, 1 H, C₄-H), 312 (d, J = 2.5 Hz, 1 H, C₁₅-H), 75 (s, 3 H, C₁₀-CH₃), 68 (s, 1 H, C₁₃-CH₃). Anal. (C₂₂H₂₈O₃) C, H.

A mixture of 1.0 g of **7b**, 0.8 g of chloranil, and 22 ml of *tert*-amyl alcohol was stirred and heated under reflux for 6.5 h.^{12b,13} After the solvent was removed by distillation under reduced pressure, the residue was dissolved in CH₂Cl₂. The CH₂Cl₂ solution was washed successively with H₂O, 5% NaOH, and H₂O again, dried (Na₂SO₄), and distilled to dryness under reduced pressure. The crystalline residue was crystallized from EtOAc to afford **7a**: mp 220–222 °C; λ_{max} ^{MeOH} 280 nm (ϵ 23750); NMR (Hz) 377 (d, J = 2.0 Hz, 2 H, C₆,C₇-H's), 344 (s, 1 H, C₄-H), 316 (d, J = 2.5 Hz, 1 H, C₁₅-H), 71 (s, 6 H, C₁₀, C₁₃-CH₃'s). Anal. (C₂₂H₂₆O₃) C, H. The NMR spectrum was identical with that of the product obtained from 1e.

3-[17β-Hydroxy-3-oxoandrosta-4,6,8(14)-trien-17α-yl]propionic Acid γ-Lactone (8). A solution of 450 mg of 7a, 4.5 ml of 1 N KOH, and 34 ml of MeOH was heated under reflux for 2 h. The cooled reaction mixture was acidified with 6 N HCl and diluted with H₂O. The resultant precipitate was collected, washed with H₂O, and dried. Crystallization from EtOAc afforded 8: mp 198-201 °C; λ_{max}^{MeOH} 341 nm (ϵ 22 800); NMR (Hz) 399 (d, J = 10 Hz, 1 H, C₇ or C₆-H), 369 (d, J = 10 Hz, 1 H, C₆ or C₇-H), 348 (s, 1 H, C₄-H), 75 (s, 3 H, C₁₃-CH₃), 63 (s, 3 H, C₁₀-CH₃). Anal. (C₂₂H₂₆O₃) C, H.

3-(16 β ,17 β -Dihydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic Acid 16,17-Diacetonide (10). A mixture of 614 mg of 3-(3 β ,16 β ,17 β -trihydroxyandrost-5-en-17 α -yl)propionic acid 16,17-acetonide (9),⁵ 615 mg of LiBr, 615 mg of Li₂CO₃, and 10 ml of DMF was stirred at 75 ± 5 °C. To this warm solution was added dropwise over a period of 25 min a solution of 0.15 ml of Br₂ in 3.5 ml of dioxane. After the addition was complete, the reaction mixture was stirred at 75 ± 2 °C for 2 h. Then it was poured into ice H₂O. The resultant mixture was neutralized with glacial HOAc. The precipitate which formed was collected, washed with H₂O, and dried: yield, 443 mg; mp 124-126 °C; λ_{max}^{MeOH} 282-284 nm (ϵ 17 500). The crude product, 10, was not purified further but used directly in the subsequent reaction.

3-(16 β ,17 β -Dihydroxy-3-oxoandrosta-4,6-dien-17 α -yl)propionic Acid γ -Lactone (5b). A mixture of 180 mg of 10, 15 ml of MeOH, 10 ml of H₂O, and 1 ml of 6 N HCl was allowed to stand at room temperature for 15 h after which it was poured into a dilute solution of NaHCO₃. The resultant mixture was extracted with EtOAc. The EtOAc extract was washed with H₂O, dried (Na₂SO₄), and evaporated to dryness to afford a crystalline product. Crystallization from EtOAc-hexane afforded 42 mg of 5b: mp 235.5–239.5 °C; λ_{max}^{MOH} 281.5–283.5 nm (ϵ 21050); ν (KBr) 3472, 1773, 1653, 1618, 1582 cm⁻¹. Another crystallization from EtOAc raised the mp to 237.5–240 °C. Anal. (C₂₂H₂₈O₄) C, H.

3- $(17\beta$ -Hydroxy-3,16-dioxoandrosta-4,6-dien- 17α -yl)propionic Acid γ -Lactone (5a). To a solution of 0.1 mg of the 16β -hydroxyspirolactone 5b in 0.1 ml of acetone was added dropwise slightly more than the equivalent amount of Jones reagent. The reaction mixture was allowed to stand at room temperature for 10 min. The excess CrO₃ was destroyed by the dropwise addition of *i*-PrOH. The resultant mixture was diluted to 2 ml with H₂O and extracted with 5 ml of CHCl₃. The CHCl₃ extract was dried (Na₂SO₄) and evaporated to dryness. The residue was subjected to TLC and GC-MS.

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References and Notes

- (a) R. C. Tweit and C. M. Kagawa, J. Med. Chem., 7, 524 (1964);
 (b) L. J. Chinn and B. N. Desai, J. Med. Chem., 18, 268 (1975).
- (2) R. C. Tweit, E. A. Brown, S. Kraychy, S. S. Mizuba, R. D. Muir, and R. T. Nicholson, *Chem. Pharm. Bull.*, 12, 859 (1964).
- (3) A. Karim, J. Hribar, W. Aksamit, M. Doherty, and L. J. Chinn, Drug Metab. Dispos., 3, 467 (1975).
- (4) K. Nakanishi, "Infrared Absorption Spectroscopy", Holden-Day, San Francisco, Calif., 1962, p 42.
- (5) L. J. Chinn and L. M. Hofmann, J. Med. Chem., 16, 839 (1973).
- (6) H. L. Dryden, Jr., and M. Kalm, U.S. Patent 3 270 008 (1966); Chem. Abstr., 67, P73768z (1967).
- (7) C. M. Kagawa, J. A. Cella, and C. G. van Armin, Science, 126, 1015 (1957).
- (8) J. A. Cella and R. C. Tweit, J. Org. Chem., 24, 1109 (1959).
- (9) L. M. Hofmann in "Recent Advances in Renal Physiology and Pharmacology", L. G. Wesson and G. M. Fanelli, Jr., Ed., University Park Press, Baltimore, Md., 1974, p 305.
- (10) L. M. Hofmann, L. J. Chinn, H. A. Pedrera, M. I. Krupnick, and O. D. Suleymanov, J. Pharmacol. Exp. Ther., 194, 450 (1975).
- (11) J. Jiu, C. Hsu, and S. Mizuba, Dev. Ind. Microbiol., 15, 345 (1974).
- (12) (a) A. B. Turner and H. J. Ringold, J. Chem. Soc. C, 1720 (1967); (b) C. C. Beard in "Organic Reactions in Steroid Chemistry", Vol. 1, J. Fried and J. A. Edwards, Ed., Van Nostrand-Reinhold, New York, N.Y., 1972, p 265.
- (13) E. J. Agnello and G. D. Laubach, J. Am. Chem. Soc., 82, 4293 (1960).